

# The Complete Mitochondrial DNA Sequence of the Deepwater Stingray *Plesiobatis daviesi* (Wallace, 1967): Unique Features in the Mitochondrial D-loop Region

Sheng-Tai Hsiao<sup>1</sup>, Chi-Hsin Hsu<sup>2</sup>, Don-Chung Liu<sup>3</sup> and I-Shiung Chen<sup>4\*</sup>

<sup>1</sup>Marine Fisheries Division, Fisheries Research Institute

<sup>2</sup>Institute of Marine Resources, National Sun Yat-sen University

<sup>3</sup>Fisheries Research Institute

<sup>4</sup>Institute of Marine Biology, National Taiwan Ocean University

## ABSTRACT

The 17514-nucleotide mitochondrial DNA (mtDNA) sequence of the deepwater stingray (*Plesiobatis daviesi*) was determined using long PCR and primer walking methods. The deepwater stingray genome contains 37 genes, including 2 ribosomal RNAs, 22 transfer RNAs (tRNAs), and 13 protein-coding genes, which are similar to other vertebrates. A comparison of the deepwater stingray mtDNA sequence with that of seven completely sequenced chondrichthyan mtDNAs revealed an identical gene order. However, the major noncoding region of the deepwater stingray genome, a D-loop sequence between the tRNA<sup>Pro</sup> and tRNA<sup>Phe</sup>, is the longest (1830 bp) among all known available sequences of chondrichthyan species. The components of D-loop sequences are two major copies of 47-bp tandem repeats and two characteristic conserved sequence blocks in this region. Phylogenetic analyses suggest that the deepwater stingray belongs to the basal group of Myliobatoidei that forms a sister group with Hexatrygonidae.

**Key words:** mitochondrial genome, Long PCR, deepwater stingray, *Plesiobatis daviesi*

## INTRODUCTION

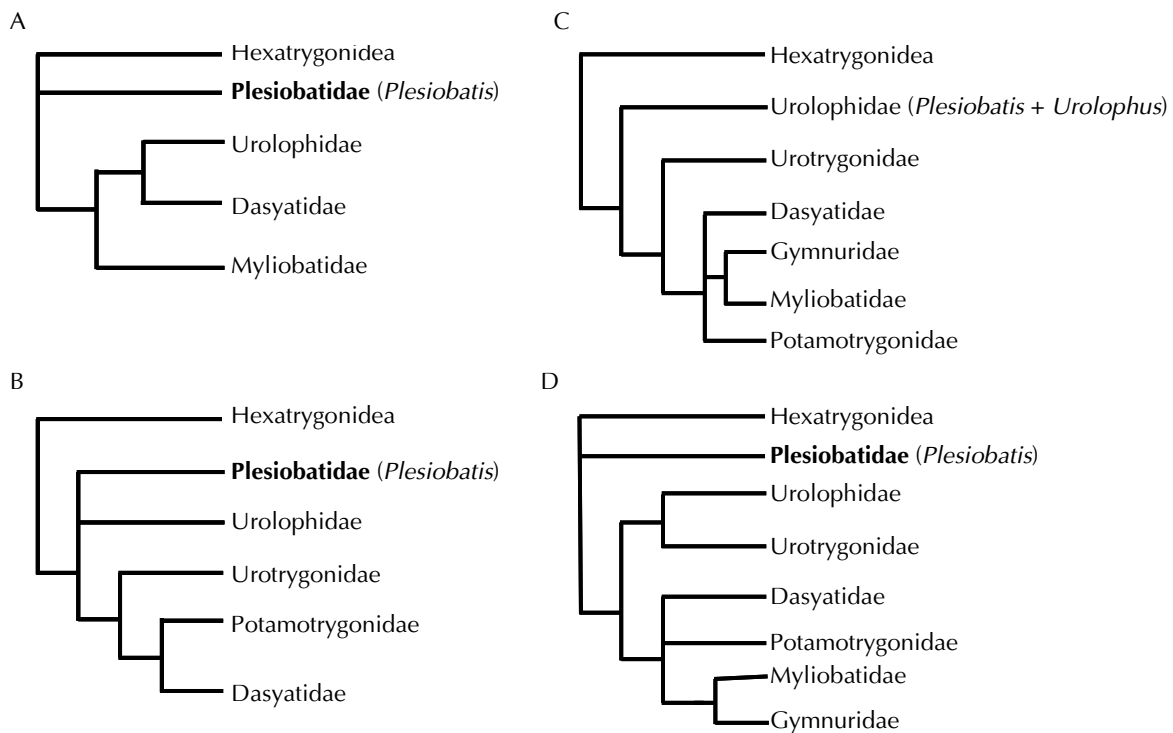
Mitochondrial genomic sequences have been used as genetic markers for molecular approaches in the phylogenetic and evolutionary studies of fish (Miya *et al.*, 2001).

The order Myliobatiformes is a monophyletic group comprised of stingrays, round rays, butterfly rays, and eagle rays. The family Plesiobatidae belongs to the order Myliobatiformes and includes only one monotypic species, *Plesiobatis daviesi*, which

was formerly recognized as *Urotrygon daviesi* and was placed under the family Urolophidae. Several different hypotheses have been proposed regarding the classification of Plesiobatidae (Fig. 1). The family was first established by Nishida (1990) based on several unique feature (i.e. nasal curtain being incompletely united and not reaching the mouth, with large size and soft disc, the average total length was ~ 200 cm and the average disc width ~ 100 cm). Later, McEachran *et al.* (1996) described musculature and skeletal structure of batoid fishes and provided a new topology on the relationship of myliobatoids. Based on the study by McEachran and Aschliman (2004) on the diversity of claspers in batoids, *P. daviesi* was placed under the family Urolophidae. However, recently Nelson (2006) placed *Plesiobatis* under the

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\*Correspondence: National Taiwan Ocean University, 2 Pei-Ning Rd., Keelung 202, Taiwan. TEL: 886-2-2462-2192 ext. 5320; FAX: 886-2-2463-3152; E-mail: isc@mail.ntou.edu.tw



**Fig. 1** The four most relevant hypotheses of phylogenetic relationship of myliobatoidei families. (A) Nishida's (1990) hypothesis based on morphological characters; (B) McEachran *et al.* (1996) proposed; (C) Phylogenetic relationship of myliobatoidei families, based on claspers characters, proposed by McEachran & Aschliman (2004); (D) The most recent hypothesis was made by Nelson (2006).

family Plesiobatidae. *Plesiobatis* is considered a primitive chondrichthyan. Therefore, in order to understand these taxa in terms of the evolutionary process among the myliobatoids, analysis of the complete mitochondrial genome sequence of the deepwater stingray would be beneficial.

Among chondrichthyans, complete mitochondrial DNA (mtDNA) sequences have been reported for only seven species: the small-spotted catshark *Scyliorhinus canicula* (Delarbre *et al.*, 1998), the star-spotted smooth-hound *Mustelus manazo* (Cao *et al.*, 1998), the horn shark *Heterodontus francisci* (Arnason *et al.*, 2001), the spiny dogfish *Squalus acanthias* (Rasmussen and Arnason, 1999a), the thorny skate *Raja radiata* (Rasmussen and Arnason, 1999b), the ocellate spot skate *Raja porosa* (Kim *et al.*, 2005), and the rabbitfish *Chimaera monstrosa* (Arnason *et al.*, 2001). In contrast, there are no published mtDNA sequences on Myliobatiformes.

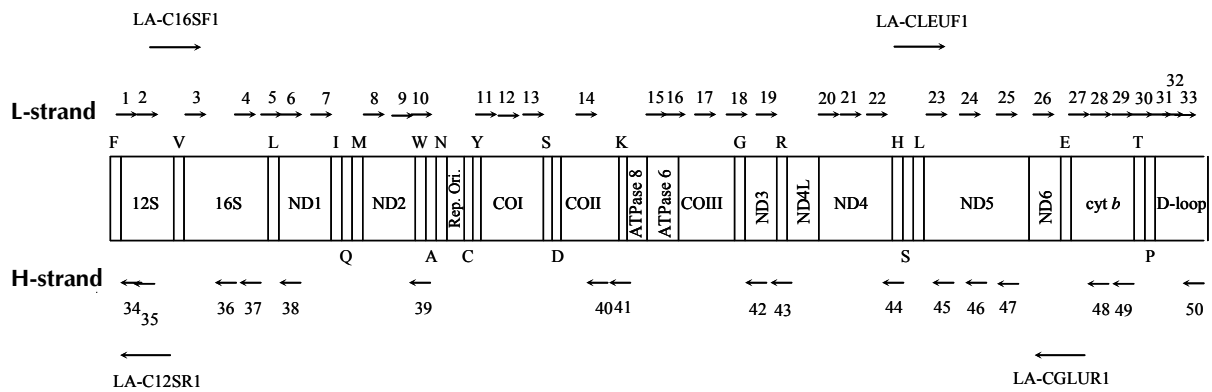
This study is to determine the complete mtDNA

sequence of the deepwater stingray *P. daviesi* (Elasmobranchii: Myliobatiformes: Plesiobatidae) was determined using long polymerase chain reaction (PCR) and primer walking methods, which have been previously applied for teleost fish (Miya and Nishida, 1999; Miya and Nishida, 2000; Inoue *et al.*, 2000, 2001a, b, c; Kawaguchi *et al.*, 2001; Miya *et al.*, 2001). This is the first study to construct the complete mtDNA sequences of myliobatoid fish may provide an independent assessment of lineage evolution within Myliobatiformes to reconcile previous conflicting hypotheses of evolutionary relationships.

## MATERIALS AND METHODS

### 1. Fish sample and DNA extraction

A deepwater stingray was captured in 2001 from Tungkang in southern Taiwan by bottom trawling at approximately 300 meters deep. Total



**Fig. 2** Gene organization and sequencing strategy for the *Plesiobatis daviesi* mitochondrial genome. The circular genome is represented linearly.

DNA extraction was performed using the protocols of the High Pure PCR Template Preparation Kit (Roche Diagnostics, Indianapolis, Ind.). The voucher specimen was deposited in the collection of the National Museum of Marine Biology & Aquarium, Pingtung, Taiwan.

## 2. mtDNA amplification by long PCR

The complete mitochondrial genome of the deepwater stingray was amplified using a long PCR technique (Miya and Nishida, 1999, 2000). Two sets of primer pairs (first set, LA-C16SF1: GCCAACCCACCTCTGTAGCAAAAAGAGAGGGAAGACTCC; LA-CGLUR1: GCTAGGGCTAGTAATTTCTGCTGGGTGGGTTGTGGTT; and second set, LA-CLEUF1: TTTCTCCCGCCCATGGTTCGAATCCC TGGCTCCCTTA; LA-C12SR1: CTCGTATAACC GCGGTGGCTGGCAGATTGACCAAC) were used to amplify the complete mitochondrial genome in two long PCR reactions.

Long PCR was performed in a Model 2400 or 9700 thermal cycler (Perkin-Elmer, Foster City, CA, USA), and reactions were performed with 30 cycles and a 25- $\mu$ L reaction volume containing 15.25  $\mu$ L of sterile distilled water, 2.5  $\mu$ L of 10 $\times$  LA PCR buffer II (Takara, Shiga, Japan), 4.0  $\mu$ L of dNTPs (0.25 mM each), 1.0  $\mu$ L of each primer (5 mM), 0.5  $\mu$ L of 1.25-unit LA Taq (Takara), and 1.0  $\mu$ L of template containing approximately 5 ng of DNA. The thermal cycle profile was that of “shuttle PCR,” that is, denaturation at 98 $^{\circ}$ C for 10 s and annealing and

extension combined at the same temperature (68 $^{\circ}$ C) for 16 min. Long PCR products were electrophoresed on a 1.0% agarose gel and then stained with ethidium bromide for band characterization via ultraviolet transillumination. The long PCR products were diluted with Tris-EDTA buffer for subsequent use as PCR templates.

## 3. Primer walking and sequencing

Fifty primers were used to amplify the contiguous and overlapping segments of the complete genome (Fig. 2). These primers were designed either from the obtained sequences of this species or from the aligned, complete nucleotide sequences of the mitochondrial genome of six species of cartilaginous fishes.

PCR was performed in a Model 2400 or 9700 thermal cycler, and reactions were performed with 30 cycles and a 25- $\mu$ L reaction volume containing 14.4  $\mu$ L of sterile distilled water, 2.5  $\mu$ L of 10 $\times$  PCR buffer (Perkin-Elmer), 2.0  $\mu$ L of dNTPs (2.5 mM each), 2.5  $\mu$ L of each primer (5  $\mu$ M), 0.1  $\mu$ L of 0.5-unit Ex Taq (Takara), and 1.0  $\mu$ L of template. The thermal cycle profile was as follows: denaturation at 94 $^{\circ}$ C for 15 s, annealing at 47–53 $^{\circ}$ C for 15 s, and extension at 72 $^{\circ}$ C for 30–60 s. PCR products were electrophoresed on a 1.0% agarose gel and then stained with ethidium bromide for band characterization via ultraviolet transillumination.

Double-stranded PCR products were purified using a High Pure PCR Product Purification Kit and

were subsequently used for direct cycle sequencing with dye-labeled (BIG-DYE) terminators (ABI Perkin-Elmer). The primers used were the same as those for PCR. All sequencing reactions were

performed according to the manufacturer's instructions. Labeled fragments were analyzed using a Model ABI 377 DNA Automated Sequencer (ABI Perkin-Elmer).

**Table 1** Polymerase chain reaction (PCR) and sequencing primers in the analysis of the *Plesiobatis daviesi* mitochondrial genome

| Primers for L-strand       |              |   |                                  |
|----------------------------|--------------|---|----------------------------------|
| Name                       | Position     | Sequence (5'-3')                              |                                  |
| Long PCR primers           |              |   |                                  |
| LA-C16SF1                  | L1417        | GCCAA CCCAC CTCTG TAGCA AAAGA GAGGG AAGAC TCC |                                  |
| LA-CLEUF1                  | L11856       | TTTCT CCCGC CCATG GTTCG AATCC CTGGC TCCCT TA  |                                  |
| PCR and sequencing primers |              |   |                                  |
| 1                          | SK-12SF2     | L199  | TAGCC CAAGA CACCT TGCTA AG       |
| 2                          | P10AN        | L501  | CAAAC TGGGA TTAGA TACCC CA       |
| 3                          | SY164F       | L1246   | TACCG CAAGG GAAAG TTGAA A        |
| 4                          | SKH-16SF3    | L2536   | AAGGG TTTAC GACCT CGATG TT       |
| 5                          | SK-LEU-UURF1 | L2815   | CTGGT AADTG CAAA GACCT AA        |
| 6                          | SK-ND1F1     | L3285   | GCMCT ACGNG CTGTA GCACA AAC      |
| 7                          | SK-ND1F2     | L3793   | TCCTA TGACA CATCA CCCTA CC       |
| 8                          | SKH-ND1F4    | L4400   | ACATT TCTGA CTACC AGAAG TACTT    |
| 9                          | SKH-ND2F2    | L4900   | TAGCC ACCAT TATAG CTCTA ACAAC    |
| 10                         | SKH-ND2F3    | L5137   | AAGCC TTCAA AGCTT TAAAC AA       |
| 11                         | SKH-ND2F4    | L5659   | ATCTA TAATG TGCTC GTAAC CGC      |
| 12                         | SKH-ND2F5    | L6142   | AATCT TAACA CAACT TTCTT TGACC    |
| 13                         | SKH-ND2F6    | L6572   | TCCTA GCCAA CTCAT CCCTC          |
| 14                         | SKH-CO2F3    | L7670   | GAAGA TGTCC TCCAC GCATG          |
| 15                         | SKH-CO2F4    | L8134   | ACTTC TTCAA TCAAT TCTTA AGCC     |
| 16                         | SKH-CO2F5    | L8559   | GAAAC TATTA GTCTA CTTAT TCGCC C  |
| 17                         | SKH-CO2F6    | L9042   | TGCGA TACGG AATAA TCTTG T        |
| 18                         | SK-GLYF1     | L9632   | AYTTA ATCTT GGTTA RAAYC CAAGG    |
| 19                         | SKH-ND3F1    | L9963   | TTCAG GCCTA ATTTA TGAAT GAC      |
| 20                         | SKH-ND3F2    | L10572  | ACTCA CATT A CCTG TTGAC TTCTC C  |
| 21                         | SKH-ND4F1    | L11088  | ACTAG GTGGT TATGG AATAA TACG     |
| 22                         | SKH-ND4F2    | L11640  | CCACC TCACC TTAAT TAACC CCT      |
| 23                         | SKH-HISF1    | L12063  | CATCA ACTCA CATCA CAACT GTAA     |
| 24                         | SK-ND5F1     | L13173  | CYCA YTAAA CGCCT GAGCC CT        |
| 25                         | SKH-ND5F2    | L13512  | ACCCA AACCT ATCCT ACCAT AAC      |
| 26                         | SKH-ND6F1    | L13930  | AACCA TGCTC ATTTT TATAG ACTCC    |
| 27                         | SKH-CYBMF6   | L14797  | AAATA TCATT CTGAG GCGCA AC       |
| 28                         | SK-LEU-CUNF1 | L15196  | CAGAG TGGTK ATGTG GTTGR CTTGA AA |
| 29                         | SK-CYBMF1    | L15213  | TTYGC CTATG CCATY YTACG CTC      |
| 30                         | SKH-DL1      | L15635  | ACCCT TATCC TTGGC TCCCA A        |
| 31                         | SKH-CYBMF5   | L15810  | AATAC ATAAG ACAGT CTATG CTAA TCC |
| 32                         | SKH-DL3      | L15972  | CCACA CTAAC AGATT CTATA CCCTA AT |
| 33                         | SKH-DL4      | L16482  | ACACC TTTGT TCGAT TTGTG G        |

**Table 1** Continued

| Primers for H-strand       |          |   |
|----------------------------|----------|---|
| Name                       | Position | Sequence (5'-3')                              |
| Long PCR primers           |          |   |
| LA-C12SR1                  | H298     | CTCGT ATAAC CGCGG TGGCT GGCAC GAGAT TGACC AAC |
| LA-CGLUR1                  | H14184   | GCTAG GGCTA GTAAT TTCTG CTGGG GTGGG TTGTG GTT |
| PCR and sequencing primers |          |   |
| 34 SK-12SR1                | H309     | CGTAT AACCG CGGTG GCTGG CACGA                 |
| 35 SK-16SR3                | H574     | TTTGA GCTAG CGCTT GTAGT ACTC                  |
| 36 SKH-16SR2               | H2059    | GCAGG CGGGA CCTCT TATA C                      |
| 37 SY163R                  | H2620    | CTCCG GTCTG AACTC AGATC ACGTA GG              |
| 38 SKH-16SR4               | H2979    | GACCT TTGCG GGATT GTATG TA                    |
| 39 SK-TRPR2                | H5132    | AAAGC TTTGA AGGCT TTTGG T                     |
| 40 SKH-CO2R4               | H7457    | GGTCA TTAAT TTCGT CCATT AA                    |
| 41 SK-LYSR2                | H7914    | TTAGC TTAAA AGGCT AATGC T                     |
| 42 SKH-ND3R5               | H9774    | GGTCA AAGCC GCACT CATAG                       |
| 43 SK-ARGR2                | H10042   | ACCAT AATTT ACTAA GCCGA AAT                   |
| 44 SKH-CO2R1               | H11802   | GCTAA TGCTG GACTG TTTAG CTT                   |
| 45 SKH-ND5R4               | H12978   | ACCTG AGCAG AGGAA GAGTA TTG                   |
| 46 SK-ND5R2                | H13331   | CTTCC GTAGG CGAGT CGTTT                       |
| 47 SK-ND5R3                | H13646   | CTTTT WGGKC CWATT TTTTC                       |
| 48 SKH-CYBMR3              | H14605   | CGGAT TATTC AGCCG TAGTT TA                    |
| 49 SKH-CYBMR2              | H14925   | GGGAG GAGGA AGTGG AATGT                       |
| 50 SKH-12SR2               | H17368   | ACATG TGTGT TTGAC ACTAT GTATG                 |

L and H refer to the light and heavy strand, respectively.

The relative positions of primers in the mitochondrial genome are shown in Fig. 2.

Posotions with mixed bases are labeled with their IUB codes.

#### 4. Sequence analysis

DNA sequences were aligned using the available complete mtDNA sequences of other cartilaginous fishes. The locations of the 13 protein-coding genes were determined by comparing the nucleotide or amino acid sequences of bony fish mitochondrial genomes. Twenty-two tRNA genes were identified by their proposed cloverleaf secondary structures and anticodon sequences, while two ribosomal RNA (rRNA) genes were identified by sequence homology and proposed secondary structures. Sequence data are available from GenBank under the accession number AY597334.

#### 5. Phylogenetic analysis

Phylogenetic relationships among

Myliobatiformes species were based on the nucleotide sequences of the mitochondrial 12S rRNA dataset from the 13 species of Myliobatiformes (e.g., Kao *et al.*, unpublished data) listed in the Appendix. DNA sequences were aligned using Clustal 1.83 as implemented in Mega 3.1 (Kumar *et al.*, 2004), and then the large gaps were manually deleted to optimize the alignment. Several approaches were applied to visualize 12S sequence diversity among the Myliobatiformes species investigated. Tree reconstruction was based on several approaches including neighbor-joining (NJ), maximum parsimony (MP), and Bayesian methods with bootstrap support where appropriate (Nei and Kumar, 2000). NJ and MP trees were constructed using Kimura's two-parameter method (Kimura, 1980) and a heuristic search with 1000 random sequence additions using PAUP\* 4.0

b10 (Swofford, 2002). Bayesian analysis was performed using MrBayes version 3.1 (Ronquist and Huelsenbeck, 2003). The best-fit model of nucleotide substitution was the GTR model, which was selected using ModelTest version 3.7 (Posada and Crandall, 1998). The selected model was GTR + I + G ( $-\ln L = 4905.52$ ,  $K = 10$ ,  $AIC = 9831.04$ ) with base frequencies of  $A = 0.3538$ ,  $C = 0.2235$ ,  $G = 0.1751$ , and  $T = 0.2476$ ; a proportion of invariable sites of  $I = 0.1777$ ; and a gamma distribution shape parameter of variable sites of  $G = 0.3799$ . The analysis was run with the best-fit model for  $1 \times 10^6$  generations, with a sampling frequency of 100 generations. The phylogenetic trees were visualized and edited with TreeView software (Page, 1996).

## RESULTS AND DISCUSSION

The mitochondrial genomic content and the base composition of each genes are listed in Table 1. The genomic content of the deepwater stingray included 2 rRNA, 22 tRNA, and 13 protein-coding genes, an origin of replication, and a control region, as found in other cartilaginous fishes (Figs 2, 3; Table 1). Its gene order was also identical to that of the other cartilaginous fishes. The arrangements of most genes were encoded on the heavy (H)-strand, excluding ND6, origin of replication, and eight tRNA genes.

With a total length of 17514 bp, the deep water stingray genome is within the range of the known genomes of other cartilaginous fishes (16697–18580 bp; Cao *et al.*, 1998; Delarbre *et al.*, 1998; Rasmussen and Arnason, 1999a, b; Arnason *et al.*, 2001). However, the total length of the deepwater stingray genome was approximately 730 bp longer than that of the thorny skate (16783 bp), which is under the order Rajiformes. By comparing these two sequences, it was found that the difference was apparently due to the length of the control region (1830 bp) of the deepwater stingray genome, which is longer than that of any other known sequences among cartilaginous fishes. However, the length of the control region of the deepwater stingray genome

was shorter than that of the rabbitfish *Chimaera monstrosa*, which is split into two control regions: NC1 and NC2 (Arnason *et al.*, 2001). Excluding the control region, the length of most genes in the mitochondrial genome of the deepwater stingray was pretty similar to that of other cartilaginous fishes (Table 2). These results indicated that the length of the coding region is somewhat conserved among teleost and cartilaginous fishes.

### 1. Protein-coding genes

Among the 13 protein-coding genes, there were four reading frame overlaps on the same strand (ATPase 8 and 6 shared 10 bp; ATPase 6 and COIII shared 1 bp; ND3 and tRNA<sup>Arg</sup> shared 2 bp; ND4L and ND4 shared 7 bp) was observed. Conversely, there was one reading frame overlap on the opposite strand (ND5 and ND6 shared 4 bp). All mitochondrial protein-coding regions began with the ATG start codon excluding COI, which starts with GTG (Table 1). The open reading frame of the deepwater stingray ended with TAA (ND1, ND2, COI, ATPase 8, ATPase 6, COIII, ND4L, ND5, ND6, and *cyt b*), TAG (ND3), and the remaining genes had incomplete stop codons of T (COII and ND4).

### 2. tRNA genes

The mitochondrial genome of the deepwater stingray contained 22 tRNA genes dispersed between the RNA and protein-coding genes (Figs. 2, 3). Its size ranged from 67 to 75 nucleotides (Table 1), which is sufficient to permit the genes to fold into the four-arm cloverleaf secondary structure. However, the tRNA<sup>Trp</sup> gene in *P. daviesi* exhibits unorthodox structures. The D-arms cannot form into a stable stem of 3–4 bp (replaced by a shorter stem of 2 bp), and the TΨC arm had 6 bp instead of 4–5 bp. There was conservation of the aminoacyl stem (7 bp) among the tRNA genes excluding tRNA<sup>Asp</sup>, which forms a stem with only 6 bp. All postulated cloverleaf structures contained 4–5 bp in the anticodon stem. As shown in Table 3, most of the tRNA genes were located on the H-strand, with the exception of tRNA-Gln, -Ala, -Asn, -Cys, -Tyr, -Ser (UCN), -Glu, and -Pro, which are located on the light (L)-strand.

| tRNA-Phe → | 12S rRNA →  
**GCTAGTGTAGCTTAAACCTAGAGCATGGCACTGAAGATGCCAAGATAAAAAATTAATCTTTTTCACAAGCA**CGAAGGTTTGGTCTAGCCTCAATATTAAT 100  
 TTTAACCTAATTTACACATGCAAGTCTCAGCACTCCGGTGAGAAGCCCTAATTAATACCTCATTATTTAGGAGCTGGTATCAGGCACATTCCAATA 200  
 GCCCATGACACCTCGCCAGCCACACCACAAGGGAAGTCAAGCAGTGTATAAACATTTGCCATAAAGCGTAAGCTTGACCAATTAAGTTATATAGTGT 300  
 GGTCAATCTCGTGCCAGCCACCGCGTTATACGAGCAACACAAATTAATTTACAGGCATTAAGGGTGTATAGAACATCTCTAATAAAAAAAGTTAA 400  
 AACCTTATTAAGCTGTACACGCTTTTATATTTAAAAACCCACTCACGAAAGTAACCTTTAAATAAATACAGAACCTTTTGACCTCAGCAGGTTAAGACC 500  
 CAAACTAGGATTAGATACCTACTATGCTTAACCAATAAACATTTGTTATAATAAACCCACCTTAATACCCGCGGAGTACTACAAGCGCTAGTCAAAACC 600  
 CAAAGGACTTGGCGGTGCTCCAAACCCCTAGAGGAGCCTGTTCTATAACCGATAATCCGCGTTCAATCCACCACCTTCTGCCTTACCACCGCCTATA 700  
 TACCGCGTGTACGCTCACCTCTCAGGGCATAAAGTAAGCAAAATGACCTTCCCTCAATACGTCAGGTCGAGGTGTAGCGAATGAAGTGGGAAGA 800  
 AATGGGCTACATCCCTTTTACAGGTATACGAACAGAAGCATGAAATCTTCTTAAGGTGATTTAGCAGTAAGTAAATTTACAGGACATATACTGAAA 900  
 CTGGCTGTAGAGCGCACACACCGCCCTCACTTCCCTCAAAATCATATTTAACTTTTAAAAAAACTTTTGTAGCAAGGAGGCAAGCTGTAACT 1000  
 GGTAAGTGTACTGAAAGTGCACCTTGGATTACCAACCAAAGTGTAGCTAAATCAGTAAAGCACCTCCCTTACACCGAGAAGATTCCCGTGCATCCGGGT 1100  
 | 16S rRNA → | tRNA-Val →  
**CACTTTGGT**TACCTCAAAGCTAGCCAAAATAAATTTATTAAGTTCCCCAATTAACCTAACACACACAACCTCTGTCTCTAATTAACATTTTTCCCTT 1200  
 CCTAGTATGGCGATAGAACAGAAACCTTTGAGCCATAGAACAGTACCAGGAAAGCTGAAAAAGAAATGAAAAACCATTAAGTAAAAAAGAACG 1300  
 AGAGACCCGCGCTCGTACCTTTGCATCATGATTTAGCAAGAACAAGTACTAGGCTAAAAAGCTTTTCTAGCCTAGCCTCCGAAACTAAACGAGCTACTTCG 1400  
 GAGCAGCTTACCCAGAGCCCAACCCACTCTGTAGCAAAAGAGAGGGAAGACTCCCAAGTACGCGGTGATAAGCTACCGAGTTTAGTGATAGCTGGTCAATC 1500  
 CAAAAAAGAAGCTTAAATCTGCATTAATTTTCAACCAGCAACTAAAAACCTTTACTAAGCTCACTTGTAATAAATTAAGAGTTATTCAAAAAAGGTAC 1600  
 AGCCTTTTGAATCAAGAAACAACCTTATAGGAGGGTAATGATTACATCTTAAAGGGTTTCTCCTCAGTGGGCTTAAAGCAGCCACTGTTAAGCAA 1700  
 GCGTCATAGCTCAAGCCTCACCTGCCCAACAATTCCTACTCATTCTCAACCCCTACTCCTACTATTGGATTATTTTAACTAATTTATAAAAAGA 1800  
 AATTATGCTAAAAATGAGTAAAGGGAATAACCCCTCCCGCACCCAGTGTATGTCAGAAAGAATTAATCACTGACAATTAACCGATGCCATAGTTGA 1900  
 GGCCTCATACATAAACAACAACCAAGAAAACCCCATACAAAACCTCGTTAACCTACACAGGAGTGTCCCGGGAAGATTAAGAAAATAAAGG 2000  
 AACTCGCAACACAACACTCCGCTTTTACAAAAACATCGCCTTGCCTCATGTATAAGAGTCCGCTGCCCTGTGATTTTTAACGGCCG 2100  
 CGGTATTCTGACCGTGCAGAGGTAGCGTAATCACTTGTCTTTAATGAAAGCCCGTATGAAAGGCATCACGAGAGTTTATCTGTCTTATTTTCCAAT 2200  
 AATGAAATGAACCTCTCGTGCAGAGCGAGAATAAACAATAAGCAGAAAGACCTATGGAGCTTCAAACACTTAAGTACTTTAAAAACATAAAAAT 2300  
 CCACTCTGGGTATAGACTAAAAACTAATTTCTTAACCTAAGCTGTTTGGTGTGGGGCAGCCAGGGGAAAAACAACCCCTTATCGAATGTGTTA 2400  
 AACAAAAAATTAGGACTACAGTCTAATCAATAGAAAATCAACGAACAATGACCCAGGGCCGATCCCTGATCAATGAACCAAGTTACCTTAGGGATA 2500  
 ACAGCGCAATCTTTCTTAGAGCCCCCATCGCGAAAGGGTTACGACCTCGATGTTGGATCAGGACATCTAATGGTGTAGCCGCTATTAAGGGCTCGT 2600  
 TTGTTCAACGATTAACAGTCTACGTGATCTGAGTTCAGACCGGAGTAATCCAGGTCAGTTTCTATCTATGATGGTATTTTCCCCAGTACGAAAGGCCG 2700  
 GGAAAAATGAAGCCTATGCCCAAAGTACGCTTCCCCCAACCTGTGAAACCAACTCAAGCAGATAAAGGCAGCCACCTTTAAACCAAGATAACGGTT 2800  
 | tRNA-Leu(UUR) → | NDI →  
**TGGGTGGCAGAGCCTGGTAAATGCGAAAGACTTAAGTCTTAAATCCAGAGGTTCAACTCCTCTCCTTAACT**ATGTTAAACTTACCCTTCCCTACATC 2900  
 M L N F T L P Y I  
 ATCAACCCCTAGCCTTCATCATCCCCATCCTTCTAGCCACAGCCTTCTTACCTTAATGAACGAAAAATTCAGGTTACATACAATCCCGAAAGGTC 3000  
 I N P L A F I I P I L L A T A F L T L I E R K I L G Y S S R K G P  
 CAAATGTAGTTGGCCGTACCGTCTTCTACAACCCATCGCTGATGGACTTAACTATTCACAAAGAACCCTACGACCTCCTTCTCTCCCAATCTCT 3100  
 N V V G P Y G L L Q P I A D G L K L F T K E P L R P S F S S Q F L  
 CTTTCTAACCCGCTCCCTCGCCTTAAACCTCGCCATTAATATGAAATCCCTACCCCTCCCACTACTCAATCCTAAACCTAAACCTAGGACTACT 3200  
 F L T A P A L L A L T L A L T L L M W M P L P L P L N P Y S I L N L G L L  
 TTTATCTAGCTATCTCAAGCCTAACAGTCTACACAATTTAGGCTCAGGCTGAGCATCAAACCTAAATACGCTCTCATGGGAGCTCCCGTGCAGTCTG 3300  
 F I L A I S S L T V Y T I L G S G W A S N S K Y A L M G A L R A V A  
 CACAACCACTCTTACGAGGTAACACTCGCCCTAATCTCTTAGCCCTACCATCTCGTAGGGGGTTTACACTCCACACATTTAACATGAGTCAAGA 3400  
 Q T I S Y E V T L A L I L L A L T I F V G G F T L H T F N M S Q E  
 AACCAATTTGACTGATCGTCCCACTGACCTTAGCCATAATATGATATATGAACTACCAAAAAACAACCCGAGCCCTTTGACCTCACAGAGGGG 3500  
 T I W L I V P T W P L A M M W Y I W T L P K T N R A P F D L T E G  
 GAATCCGAAGTGTCTCAGGTTTCAACACTGAGTACGAGGGGGCTCTTTCGGCCTATTCTTCTAGCCGAATACCTCAATATCTTACTAATAAATACC 3600  
 E S E L V S T G F N T E Y A G G S F A L F F L A E Y S N I L L M N T L  
 TCTCGCTATCTTCTAGGCGCTCTACACCCACACTTCCCAACACTAGTACCCTTAACTTACCTCAAAAACAACCCGACTAACCTACTACT 3700  
 S A I L F L G A S Y T P H F P Q L A C T T L N L M L K T T A L T L L F  
 TCTATGAATTCGAGCTCTTACCCGATTCGATACGACCAACTCATACACTTGTCTGAAAAAATCTTACCAATAACACTAGCCCTGATCCTATGA 3800  
 L W I R A S T Y P R F R Y D Q L M H L V W K N F L P M T L A L I L W  
 CACATCACCTACCAATCATTACAGCAAGCCTCCACCAATAAATAACTTAAAGCGAAAGTGCCTGAATAAAGGGCCACTTTGATAGAGTGGATAACGAA 3900  
 H I T L P I I T A S L P P M M T \*  
 | tRNA-Gln → | tRNA-Ile →  
**TGTAAAACCATTCCTTTCTCT**CTAGAAAGACAGGATTTGAACCTATACCTAGAGATCAAAACTCTTTGTACTCCCACTATACTACTTCTCTA 4000  
 | tRNA-Met → | ND2 →  
**AAAGTCAGCTAAATCAAGCTTTTGGGCCCA**TACCCCAACATGTTGGTTAAATCCTTCTCTTACTAATGAACCCATTAATCTCACTATCTCAATCCTT 4100  
 M N P L I L T I S I L  
 AGTCTAGGATTAGGCACCACAATAACACTCATTACCTCACACTGATTACTAATCTGACTTGGCCTCGAAATTAATACAATAGCCATATCCCCCTCATAA 4200  
 S L G L G T T M T L I T S H W L L I W L G L E I N T M A I I P L M I  
 TCCATCAACACCATCCCGTGCAAGCAAGCAACTACAAAATCTTCTTACACAAGCCACTGCCCTCAGCCCTCCTCTTATTTGGCAGCACCACCAACGC 4300  
 H Q H H P R A T E A T T K Y F L T Q A T A S A L L L F A S T T N A  
 CTGGCTACCCGGCCAATGAAATGACCGAAATAATTAATCCCTTCCACCACTTCTCTCCATTGGCTTAGCACTAAAAATGGCTTAGCCCTTAA 4400  
 W L T G Q W N M T E M I N P L S T T L S I A L A L K I G L A P L  
 CATTCTGACTACCAAGTACTTCAAGGACTTAACTTACTCACAGGACTCATCTCTATCATGACAAAAACTTGCACCATTTCGCAATTTACTACAAC 4500  
 H F W L P E V L Q G L N L L T G L I L S T W Q K L A P F A I L L Q L  
 TCTACCCCTCTTAAACCAACATTACTTGTATCAATGGGCATACTATCAATCGTCATCGGTGGTTGAGGCGGCCCAACCAACACAACCTACGAAAAAT 4600  
 Y P L L N P T L L V S M G M L S I V I G W G G L N Q T Q L R K I  
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 L A Y S S I A H I G W M T V I L H Y S P N L T L L N L I I Y I I M  
 ACCTCCTCATTATCTCTCTTCAACAACACACCAAAAATAATTAATCAATCTCCACATCATCAACTAAATCCCACTACTATCCATTAACCA 4800  
 T S S L F L L Y N N T T K I N S I S T S S T K S P L L S I M T M  
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 L T L L S L G G L P P L T G F M P K W L I L Q E M T K Q N L L I L  
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 A T I M A L T T L L S L F F Y L R L C Y M I S L T L P P N P I S S  
 TCATCATGACGAACAAAACCTTCAACCCAACTAACCTTAACTCCCTCTTCTCCTCCATTGACCCCAACTACTTCACTAA 5100  
 S S S W R T K T I Q P N L T L M T S L S L L L L P L T P T M L S L T  
 CTACTACATAAAGAAATTTAGGTTCAAATAAACCAAAAGCCTTCAAGCTTTAAACAAGAGTGAAAACCCCTTAATTTCTCTAAGGTTTGCAAAACCTT 5200  
 T T \*  
 | tRNA-Trp → | tRNA-Ala →  
**ACTTCACATCTCTTGAATGCA**ACCCAAAGTCTTTAATTAAGCTAAACCTTACTAGATAAAACAGGCCCTCGATCCTATAAAATCTTAGTTAACAGCTAAGT 5300  
 | Rep-Origin → | tRNA-Cys →

GTCCAATCCAGCGAAGCTTTTATCTAGCTTCTCCCGCGTTTCAGGGCGGGAGGGCGGGAAGCCCGGGGAGAAGCCTATTCTCCTTCTCGGGACTTGCAA 5400  
 + tRNA-Tyr →  
 TCCCGTGTATTATAACTACAGGGCTGAC TAGTAAGAGGAGGACTCGAACCTCCCTCTACGGGGCTACAATCCCGCGCTTTGGGTCTCAGCCATCTTAC 5500  
 + tCOI →  
 IGTGACAATTAATCGTTGATTATTTCTACCAATCACAAGACATTGGCACCCTTATTTAATCTTTGGTGCATGGGCAGGAATAGTGGGTACCGGCCT 5600  
 V T I N R W L F S T N H K D I G T L Y L I F G A W A G M V G T G L  
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 S L L I R T E L S Q P G A L L G D D Q I Y N V L V T A H A F W M I  
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 F F M V M P I M I G F G F G N W L V P L M I G A P D M A F P R M N N M  
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 S F W L L R T P P S F L L L A S A G V E A G A G T G W T V Y P P L A  
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 G N L A H A G A S V D L A I F S L H L A G A S S I M A S I N F I T  
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 T I I N M K P P A I S Q Y Q T P L F V W S I L I T T V L L L L S L P  
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 V L A A G I T M L L T D R N L N T T F F D P A G G G D P I L Y Q H  
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 T R A Y F T S A T M I I A I P T G V K V F S W L A T L H G G T I K  
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 W E T P L L W A L G F I F L F T V G G L T G I I L A N S S L D I V  
 CTTACGATACTTACTAGTTGAGCCACTTCCATTATGTTTATCAATAGGGCAGTATTCGCTATCATAGCTGGTTTCGTCCACTGATTTCTCTTAA 6700  
 L H D T Y Y V A H F H Y V L S M G A V F A I M A G F V H W F P L I  
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 T G Y T L H P T W T K Y Q F L V M F V G V N M T F F P Q H F L G L  
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 I M M F I L W E A F A S K R E I L H I E L P H T N V E W L H G C P P  
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 P Y H T Y E E P A F V Q V Q Q P \*  
 + tRNA-Ser →  
 CAACACATAACCCTCTGTCACTTCTTCTGAGATTCTAGTAAAACCTTACATTTCTTGTCAAGGCAAAATGTGGGTTTAAACCCCAAGAACTTTAA 7200  
 + tRNA-Asp →  
 + tCOII →  
 IATGGCACATCCATCACAATTAGGTTTCAAGACGCGAGCTCTCCAGTTATGGAAGAGCTCCTCCACTTTCACGATCATACCTTAATAATCGTATTTCTC 7300  
 M A H P S Q L G F Q D A A S P V M E E L L H F H D H T L M I V F L  
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 V P A I I L V M I A V P S L R I L Y L M D E I N D P H I T I K T I  
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 G H C Q W Y W S Y E Y T D Y Q N L E F D S Y M T Q T E N L T P G Q F  
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 L G I K M D A V P G R L N Q T A F I I S R P G I F Y G Q C S E I C  
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 G A N H S F M P I V V E A V P L E H F E N W S S L M L E E L \*  
 + tRNA-Lys →  
 AGCTAAACAGTCCAGCATTAGCCTTTTAAGCTAAAATTTGGTGACTCTTAACCACCTTAATGACATGCTCAACTTAAACCCGGTCCCTGATTTTTAAT 8000  
 M P Q L N P G P W F L I  
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 F L F T W L F F L A I M P N K V M S Y L L N N N P T T K N N Q K P  
 + ATPase6 →  
 AAACCAAAACCCTGAAATTGACCAATGGCCCTAAACTCTTCAATCAATTTCTTAAGCCATCACTACTTGGACTCCCCCTATTGCTCTAGCAATCATAAT 8200  
 K P N P W N W P W P \*  
 M A L N F F N Q F L S P S L L G L P L I A L A I M I  
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 P W L I F P P P S K Q W L T N R L L T L Q T W F I N R F T H Q L I  
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 T P T T Q L S L N M A F A L P L W L T T V L I G M F N Q P T V A L  
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 R L T A N L T A G H L L M Q L I A T A A F A L I S I M P S I A L L T  
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 S L I L L F L T I L E V A V A M I Q A Y V F V L L C S L Y L Q E N  
 + tCOIII →  
 TGTTTAAATGGCCATCAAAACATGCATATACATAGTTGACCCAGCCATGACCCTTAAACAGGAGCAGTAGCAGCCCTTCTCAACCTCAGGTCTCG 8900  
 V \* M A H Q T H A Y H M V D P S P W P L T G A V A A L L M T S G L V  
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 I W F H F H S L P L L L C L G L L L T L T I Q W R D V I R E G  
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 N T A V L L A S G V T V T W A H H S V M E G N R K E A I Q A L A L



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V A T G F H G L H V M I G S S F L L V C L L R Q I Q Y H F T S Q H H  
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F G F E A A A W Y W H F V D V V W L F L Y V S I Y W W G S \*  
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M N L I T F V V A L T A L  
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I S L I L A M L A F W L P T L S P D N E K M S P Y E C G F D P L G  
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T A R L P F S L R F F L V A I L F L L F D L E I A L L P L P W A  
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V Q L D S P L I T S F W A T T I L L L L T S G L I Y E W L Q G G L E  
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W A D \*  
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T F S S A F A L S L L G L A L N R S H L L S A L I C L E G M M L S  
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L F T A I A L W S T T M T T P T C S L A P M I L L T F S A C E A S  
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A G L A L L V A T T R T H G S D M L K N L N L L Q C \*  
M L K I I I P T I  
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M L Y P I S W A T P K K W L W T A S T S Y S L L I A F I S L S W F  
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M L L A S Q N H L N E P H T R Q R I Y I N L L I T L Q L L I L  
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R L N A G I Y F L F Y T L M A S L P L L I A L L A L Q N D F G S L S  
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M L T F Q F P Q L L N S S W T N K F W W A A C L I A F L V K M P  
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L Y G T H L W L P K A H V E A P I A G S M I L A A I L L K L G G Y  
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G M M R I I S I L D P L T K E M A F P F L I L A L W G I I M T S S T  
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C L R Q T D L K S L I A Y S V T H M G L V V A A I L I T Q T P W S  
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F A G A T A L M I A H G L I S S A L F C L A N T N Y E R T H T R T  
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M G E L L I M S S L F K W S Q W T I I M T G L G V L L T A S Y S L  
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L P S L L L I S K P E L I L G W T S \*  
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N L N K F F K Y L L L F L I T M L I L I T A N N L F Q L F I G W E  
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G V G I M S F L L I G W W L S R A D A N T A A L Q A I I Y N R I G D  
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I G L I T A M A W L A M N L N S W E I Q Q L F F L S K N T D L T L  
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C L L G L V L A A A G K S A A T F G L H P W L P A A M E G P T P V S  
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A L L H S S T M V V A G I F L L I R L H P L I Q D N Q L V L S A C L  
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C L G L A T C A L T C A L T Q N D I K K I I A F S T S S Q L G  
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L M M V T I G L N Q P Q L A F L H I C T H A F K A M L F L C S G  
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S I I H S L N D E Q D I R K M G M H K L L P F T S S S L T V G S L  
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A L T G M P F L S G Y F S K D A I I E A M N T S H L N A W A L T L  
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T L L A T S F T A I Y S L R L T S F S L M N Y P R F T P L S P I N

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 YAQTQVSTQMI D L L S W N E K I G P K S P I I Q Q T F L I K L S  
 CAACCTCCCCTCAACAAGGCTTAAACAAACATACCTTCTCTACTTCTCTTACCTTACCTTATCTCAGCCCTCACCTTACTTAAACCGTCCGCAA 13800  
 T S P Q Q G L I K T Y L L L L L L T L T L S S A L T L L \* V T R L  
 AGCCCTCAAGATATCCCTCGCGTCAACTCAAACACCACAAACAAAGTCAAAGCAACATTCAACCCCTAAAACTAAAACCTCACTCCCTCAAGAATAC 13900  
 AGWSMGRTELVVFLTL LLMWGGGLV L V G S G W S Y  
 AATAAGCCACCCCACTGAAATCCCAACCAACCATGCTTCTTATAGACTCTCACCCTCCCACTCAACCCCTCAACTCAATCAACCACAAAATTA 14000  
 L L A V G S F D G R V M S M E M S E E G G V W G V W D V F N  
 AACCCACTCCAAAAAGAAATCCAAAATACCAACACATAAGCTAAAACAGACCAATCACCCAGACTCAGGATAAGCTCAGCAGTAAACCGCCGT 14100  
 L G V G F L I G F Y G V Y A L V S W D G W S E P Y P E A A L A A T  
 ATAAAGCAACACAATATTCACCCCAAAATAAAACAAAATCAAAGACAAAAAAGAACTCCAAATCAACCAATAAACCAACCAACCCCA 14200  
 Y A F V M M M G G L Y I L F L I L S L F S S G F G V L L G C G V G  
 GCAGAAATACCTAGCCTAGCGCAAAAATAGGAGAAGGATTCGATGCCACAGCTACTAAGCCATAAACCAAACTCACTTTAAATCAACATTACAT 14300  
 A S I V L G L A A F Y P S P N S A V A V L G L V F S V M L I L M V  
 ←ND6 + tRNA-Glu→ + cyt b→  
 ACATCATTTATTCCTACTTAGACTCTAACTAAGACCTTTAATCCGAAAAACTACCGTTGTTATTCAACTATAGAAAACCCCTTAATGACTACTACAACACCC 14400  
 Y M M M T T T N T R  
 GCAAAACCCATCCCTTATCAAATATCAATAACTCCGTAATGACCTCCCAACTCCAATAACATCTCTGCCTGATGAACTTCGGCTCACTTGTAGG 14500  
 K T H P L F K I I N N S V I D L P T P T N I S A W N F G S L V G  
 CCTATGCTTAATTCAAAATCCTGACAGGCTCTTCTCAACTATACATACACCGCAGACATCTCAACCGCATCTCTCAGTAGCACATATCTGCCGA 14600  
 L C L I I Q I L T G L F C L M H Y T A D I S T A F I S V A H I C R  
 GACGTAACACTACGGCTGAATAATCCGCAACCTTCAGCCCAATGGCGCTCCCTCTTTTTCATCTGCGTTTACCTCCACATCGCCCGAGGACTTTACTAG 14700  
 D V N Y G W M I R N L H A N G A S L F F I C V Y L H I A R G L Y Y G  
 GCTCCTACCTAAACAAGAAACCTGAAATATTGGGTAATATCTCTTACTCCTTATAGCCACTGCCTTCGTAGGATACGTCCTCCATGAGGACAAAT 14800  
 S Y L N K E T W N I G V I I L L L M A T A F V G Y V L P W G Q M  
 ATCATCTGAGGCGCAACCGTCATCAACACTACTACCGCCCTCCCTTATATTGGAGACACACTCGTCAATGACTCTGAGGTGGCTTCTCAATCGAC 14900  
 S F W G A T V N L S A L P Y I G D D T L V Q W L W G F S I D  
 AATGCAACACTAACCGATCTTTACATTCACCTCTCTCCCTTCTAATCGGAGCCCTAACTATAGTCCACCTTCTCTCCACACCTCCGGTT 15000  
 N A T L T R F F L F H F L P F L I A A L T M V H L L F L H T S G S  
 CAAATAACCAACTGGTCTTCCCTTGACATAGACAAAATCCCATTTACCCCTACTACTATTAAAGACCTCTCGGCTTCTTCTCCTCTGCTCT 15100  
 N N P T G L P S D M D K I P F H P Y Y S F K D L L G F F L L L L L  
 ACTTACCTCTTAGCTTACTCACACCACTCTAACCAGCAGAAAACCTTTATCCAGCCAAACCTCTCGTCAACCCACACATCAAAACAGAG 15200  
 L T L L A L L T P N L L T D T E N F I P A N P L V T P P H I K P E  
 TGGTACTTCTATTCGTTACGCCATCTACGCTCTATCCCAACAACTAGGAGGCGTACTTGCCTTGCCTTTCATCTTAATCTCTTCTTAGTCC 15300  
 W Y F L F A Y A I L R S I P N K L G V L A L A L S I L I L F L V P  
 CCATTCTCATACTTAAACTACGAAGCCTAACCTTTCGCCGATCACACAGCTTCTATTCTGGCTTATAGTGGCAATACCTTATCTAACATGAAT 15400  
 I L H T S K L R S L T F R P I T Q L L F W L V A N T L I L T W I  
 CGGAGGACAACCGTAGAACAACTTATATCAATTGGCCAAATCGCTCCTACCTATTCTCTCTTCTTCCCTTTTCCAACTCGCCGGTTGA 15500  
 G G Q P V E Q P F I I I G Q I A S I T Y F S F F L L L F P I A G W  
 TGAGAAAACAAAATACTAAACCTTAAACCAATGCTCTGGTGGCCAGCTATTTAAGGCATCGGCTTGTAAACCGAAAATCGGAGGTGAAATTCCTCC 15600  
 W E N K M L N L \*  
 + tRNA-Pro→ + Control region→  
 CAGTGCATAAAGCACTTTCAGAAAAAGAGGACAAACCCCTTATCCTTGGCTCCAAAGCCAAGATTTTATTAACCTATTTCTCAACCTTATTAAGAAA 15700  
 AGTCGACTAAATATGACGTTAGTTGACTAGATCACATATAATTTAAACCCCTATAATGCTTAAATACATAATACATTAATAAATTAATCCCATATA 15800  
 Repeat 1 Repeat 2  
 TTGTCATATAATACATAAGACAGCTATGCTTAACTCTCATACATCTATATACCCCTATATCATATAATCTATGCTTAACTCTCATACATCTATATA 15900  
 Repeat 3 (partial)  
 CCCTATATCATAAACATATCTATGCTTAACTCTCATACATCTATATACCCCTATTTGATTACCCACTCAACCTCACACTAACAGATCTATACCCCTAATCT 16000  
 ATTTATCCACCAACAGCTCTCAATTTATTTAACAACAGTCCCATTAACAGATAACCTCCCAACCTATATTAACCATACGCTTAACTAACATATCAA 16100  
 TAS  
 GATAITCTATTTATTCATAGCATATATAACCTTAACTACCATAAATGACATACATACCTTCTGCTTATCAGCTAGGATTAATACCTTCTTACTTCCAGTCTCTC 16200  
 TAACTCCCACTACCGTACTGGTTAATTTCACTATTAATCATCTATTCACCTTCTGCTTATCAGCTAGGATTAATACCTTCTTACTTCCAGTCTCTC 16300  
 CAAAATCTATAACTTATCAATGTTAATCAATTAATTAICGATAATATAACTAICTATGCTTAAICAGCATTAACTCGACATCCCTTATTCATAAAT 16400  
 CSB-D  
 CAGATTCAAACATTAACACATTTCTATTTCTCCGATAATTTCTTGGTTGGCGTGAATAATCAACGATCTAATAGAATACACCTTTTGGTTCGATTTGT 16500  
 GGCACAGAAAATAGATTTATCCGATAAATGATCAGACCGACATTTGGCACCTTAACACGCAATACGCTCTTAACTCGGTCAGAACTCACTTCCCTTA 16600  
 GTTCCCTTGTAAGGCACAACCTTGTGACCCAACTCTGCTACGACGGATCGGGTCTACTAAGATTCAGCTCCGCTAAGTCTTTTTAGTTTCTGTTGA 16700  
 CTTCATCCACTATTTCTTCTGTTACGGTACTGCTGATAGGATGGTAAAGACCTCGACTACAAGTAAATGATTAAGTACTTCTTCAATCACGATGATCACT 16800  
 CGGTCCAGAGGACTACCGTTGAAATTAACAAAAGGTTATAGTTGAGAAATTAATCAATTTCTATAATTAATTTAGGCAATTAATCAAAAAAGACATGTA 16900  
 AAAATTTTTCATAAGAAATTAATTAATTAATTTGATAAATAGGAATAACAATGAGCAAGTCTTAAATCTGAAAGTAAAGTGGTCCCTCCCTGT 17000  
 CSB II CSB III  
 GCGGGCGTGGTAAAAGATATTTATACAAAGAAATTTTTGGGAAAAACCCCTCCCAAAAAACCCGGTATCTCGAAAAACCCCTAAAACGAG 17100  
 GGCAGAAAGAACTTTTTTATGCCATAAATAATCCAAAAGAAATTTTTTATTTAAAACAAAAAATTTTTTATTTAAACAACTTTTATTTAAACAACTGGCAAAACGAAATTA 17200  
 ACTTGGGGAATAAATAAGACTACTCATGATGATTTAAAAAATAAATAAAGATTAACCGTGTCCAGGACATTAATCTGGGATCTATATGTTGAAAAATTAAC 17300  
 TTTTTTTTCCCTAATATACCTGTTATATTTAAATTTTTTACTGGTTGGTGGCGTGTGTGTGATGCAATACATAGTGTCAAAACACACATGTTGATTAAC 17400  
 TGAATGTCCCTACATATACACACAAATATGCAATTTTTGACAAAAAACACACATACACACATTTTCTCCACCATATCTTAACCTTCACTTCAAACT 17500  
 CTTCACATGATC 17514

**Fig. 3** The complete L-strand nucleotide sequence of the deepwater stingray mitochondrial genome. Position 1 corresponds to the first nucleotide of the tRNA Phe gene. The direction of transcription for each gene is shown by arrows. The beginning and end of each are indicated by vertical bar (| and |). Transfer RNA genes are boxed; corresponding anticodons are indicated in black boxes. Amino acid sequences presented below the nucleotide sequence were derived using mammalian mitochondrial genetic code (one letter amino acid abbreviations placed below the first nucleotide of each codon). Stop codons are overlined in gray boxes and indicated by asterisks. Non-coding sequences are underline with dots. TAS, putative termination-associated sequence; CSB2, 3, and D, conserved sequence blocks. Sequence data are available from DDBJ/EMBL/GenBank with accession number.

**Table 2** Comparisons of lengths (bp) of 8 cartilaginous fish mitochondrial genomes

| Subclass         | Holocephali                  | Elasmobranchii                | Elasmobranchii           | Elasmobranchii                | Elasmobranchii                              | Elasmobranchii             | Elasmobranchii                              | Elasmobranchii           |
|------------------|------------------------------|-------------------------------|--------------------------|-------------------------------|---|----------------------------|---|--------------------------|
| Order            | Chimaeriformes               | Carcharhiniformes             | Carcharhiniformes        | Heterodontiformes             | Squaliformes                                | Myliobatiformes            | Rajiformes                                  | Rajiformes               |
| Family           | Chimaeridae                  | Scyliorhinidae                | Triakidae                | Heterodontidae                | Squalidae                                   | Plesiobatidae              | Rajidae                                     | Rajidae                  |
| Species name     | <i>Chimaera monstrosa</i>    | <i>Scyliorhinus canicula</i>  | <i>Mustelus manazo</i>   | <i>Heterodontus francisci</i> | <i>Squalus acanthias</i>                    | <i>Plesiobatis daviesi</i> | <i>Raja radiata</i>                         | <i>Raja porosa</i>       |
| Common name      | rabbitfish                   | small-spotted catshark        | starspotted smooth-hound | horn shark                    | spiny dogfish                               | deepwater stingray         | thorny skate                                | ocellate spot skate      |
| Accession number | AJ_310140                    | Y_16067                       | AB_015962                | AJ_310141                     | Y_18134                                     | AY_597334                  | AF_106038                                   | AY_525783                |
| 12S rRNA         | 949                          | 957                           | 952                      | 952                           | 951   | 960                        | 967   | 965                      |
| 16S rRNA         | 1665                         | 1673                          | 1669                     | 1676                          | 1676  | 1689                       | 1678  | 1676                     |
| ND1              | 972                          | 975                           | 974                      | 976                           | 975   | 978                        | 975   | 975                      |
| ND2              | 1044                         | 1047                          | 1046                     | 1047                          | 1047  | 1044                       | 1047  | 1047                     |
| COI              | 1560                         | 1554                          | 1556                     | 1557                          | 1557  | 1551                       | 1557  | 1557                     |
| COII             | 691                          | 691                           | 690                      | 691                           | 691   | 691                        | 691   | 699                      |
| ATPase 8         | 168                          | 168                           | 167                      | 168                           | 168   | 168                        | 168   | 168                      |
| ATPase 6         | 684                          | 684                           | 683                      | 684                           | 684   | 684                        | 684   | 684                      |
| COIII            | 786                          | 786                           | 785                      | 786                           | 786   | 786                        | 786   | 786                      |
| ND3              | 349                          | 351                           | 350                      | 351                           | 351   | 351                        | 351   | 351                      |
| ND4L             | 297                          | 297                           | 296                      | 297                           | 297   | 297                        | 297   | 297                      |
| ND4              | 1375                         | 1381                          | 1380                     | 1381                          | 1381  | 1387                       | 1381  | 1387                     |
| ND5              | 1839                         | 1830                          | 1826                     | 1830                          | 1833  | 1824                       | 1836  | 1836                     |
| ND6              | 522                          | 522                           | 521                      | 510                           | 522   | 522                        | 519   | 519                      |
| cyt b            | 1144                         | 1144                          | 1145                     | 1146                          | 1146  | 1146                       | 1143  | 1143                     |
| Control region   | 1602, 1503                   | 1050                          | 1067                     | 1068                          | 1080  | 1830                       | 1064  | 1310                     |
| Total            | 18580                        | 16697                         | 16707                    | 16708                         | 16738                                       | 17514                      | 16783                                       | 16972                    |
| Reference        | Arnason <i>et al.</i> (2001) | Delarbre <i>et al.</i> (1998) | Cao <i>et al.</i> (1998) | Arnason <i>et al.</i> (2001)  | Rasmussen <i>et al.</i> (1999) <sup>1</sup> | present study              | Rasmussen <i>et al.</i> (1999) <sup>2</sup> | Kim <i>et al.</i> (2005) |

### 3. Ribosomal RNA genes

The length of 12S and 16S rRNA genes of this deepwater stingray were 960 and 1689 bp, respectively (Table 1). They are located between the tRNA<sup>Phe</sup> and tRNA<sup>Leu(UUR)</sup> genes and separated by the tRNA<sup>Val</sup> gene (Figs. 2, 3).

### 4. Noncoding sequences

The L-strand replication origin in the deepwater stingray was located between the tRNA<sup>Asn</sup> and tRNA<sup>Cys</sup> genes, and it was 34 bp in length (Fig. 2), indicating that it has the potential to fold into a stem-loop secondary structure. This region has been observed in most cartilaginous fishes.

The major noncoding region found in the deepwater stingray was located between tRNA<sup>Pro</sup> and tRNA<sup>Phe</sup> and was up to 1830 bp in length, which was much longer than that of the sharks and other rays. Several unique characteristics were identified in the control region (Fig. 3), namely three conserved

sequence blocks (CSB II, III, and D; Kawaguchi *et al.*, 2001), a termination-associated sequence, and the two copies of 47-bp repeat regions (repeat 1 and 2). Repeated sequences have also been observed in teleosts (Cecconi *et al.*, 1995; Lee *et al.*, 1995; Chen *et al.*, 1998; Chen *et al.*, 2002; Stefanni *et al.*, 2002). However, only the first 18 bp of the 3<sup>rd</sup> repeat region were identical to those of the 1<sup>st</sup> and 2<sup>nd</sup> repeat regions. Tandem repeats were located near the 5' end of the control region. Such repeats may result from strand slippage of the third displaced DNA strand in the D-loop (Buroker *et al.*, 1990).

### 5. Phylogenetic relationships among other Myliobatoidei families

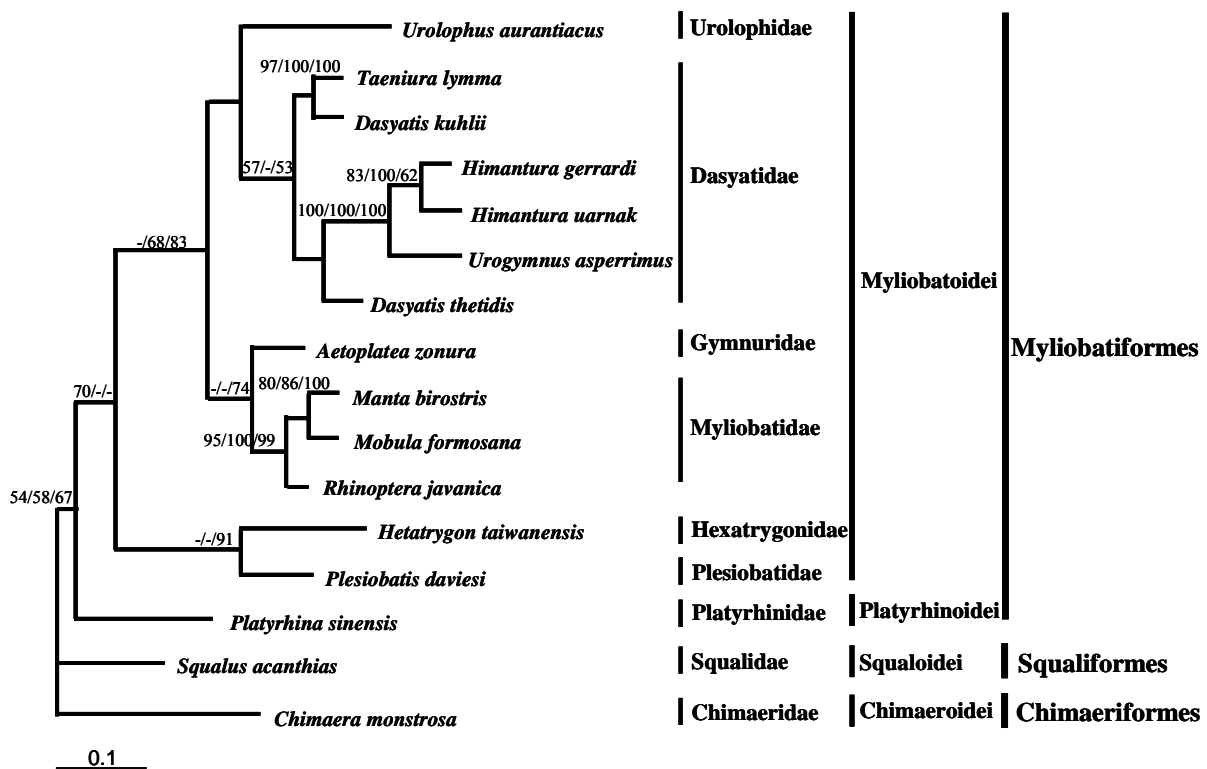
To determine the phylogenetic position among Myliobatoidei families, the 12S rRNA sequences of 13 fish species were retrieved from GenBank (Appendix). Sequence alignment resulted in 748 molecular characters. The same topology of

**Table 3** Location of features in the mitochondrial genome of *Plesiobatis daviesi*

| Gene                         | Position number |       | Size (bp) | Codon |      | Strand |
|------------------------------|-----------------|-------|-----------|-------|------|--------|
|                              | From            | To    |           | Start | Stop |        |
| 1 tRNA <sup>Phe</sup>        | 1               | 70    | 70        |       |      |        |
| 2 12S rRNA                   | 71              | 1030  | 960       |       |      |        |
| 3 tRNA <sup>Val</sup>        | 1037            | 1108  | 72        |       |      |        |
| 4 16S rRNA                   | 1110            | 2798  | 1689      |       |      |        |
| 5 tRNA <sup>Leu(UUR)</sup>   | 2799            | 2873  | 75        |       |      |        |
| 6 ND1                        | 2874            | 3851  | 978       | ATG   | TAA  |        |
| 7 tRNA <sup>Ile</sup>        | 3854            | 3922  | 69        |       |      |        |
| 8 tRNA <sup>Gln</sup>        | 3925            | 3996  | 72        |       |      | L      |
| 9 tRNA <sup>Met</sup>        | 3998            | 4067  | 70        |       |      |        |
| 10 ND2                       | 4068            | 5111  | 1044      | ATG   | TAA  |        |
| 11 tRNA <sup>Trp</sup>       | 5112            | 5181  | 70        |       |      |        |
| 12 tRNA <sup>Ala</sup>       | 5183            | 5251  | 69        |       |      | L      |
| 13 tRNA <sup>Asn</sup>       | 5253            | 5325  | 73        |       |      | L      |
| 14 Rep-Origin                | 5326            | 5359  | 34        |       |      | L      |
| 15 tRNA <sup>Cys</sup>       | 5360            | 5427  | 68        |       |      | L      |
| 16 tRNA <sup>Tyr</sup>       | 5431            | 5501  | 71        |       |      | L      |
| 17 CO I                      | 5503            | 7053  | 1551      | GTG   | TAA  |        |
| 18 tRNA <sup>Ser(LUCN)</sup> | 7059            | 7129  | 71        |       |      | L      |
| 19 tRNA <sup>Asp</sup>       | 7131            | 7197  | 67        |       |      |        |
| 20 CO II                     | 7202            | 7892  | 691       | ATG   | T--  |        |
| 21 tRNA <sup>Lys</sup>       | 7893            | 7964  | 72        |       |      |        |
| 22 ATPase 8                  | 7966            | 8133  | 168       | ATG   | TAA  |        |
| 23 ATPase 6                  | 8124            | 8807  | 685       | ATG   | TAA  |        |
| 24 CO III                    | 8807            | 9592  | 786       | ATG   | TAA  |        |
| 25 tRNA <sup>Gly</sup>       | 9593            | 9663  | 71        |       |      |        |
| 26 ND3                       | 9664            | 10014 | 351       | ATG   | TAG  |        |
| 27 tRNA <sup>Arg</sup>       | 10013           | 10084 | 72        |       |      |        |
| 28 ND4L                      | 10085           | 10381 | 297       | ATG   | TAA  |        |
| 29 ND4                       | 10375           | 11755 | 1387      | ATG   | T--  |        |
| 30 tRNA <sup>His</sup>       | 11756           | 11824 | 69        |       |      |        |
| 31 tRNA <sup>Ser(AGY)</sup>  | 11825           | 11892 | 68        |       |      |        |
| 32 tRNA <sup>Leu(CUN)</sup>  | 11894           | 11965 | 72        |       |      |        |
| 33 ND5                       | 11966           | 13789 | 1824      | ATG   | TAA  |        |
| 34 ND6                       | 13786           | 14307 | 522       | ATG   | TAA  | L      |
| 35 tRNA <sup>Glu</sup>       | 14308           | 14376 | 69        |       |      | L      |
| 36 cyt <i>b</i>              | 14382           | 15527 | 1146      | ATG   | TAA  |        |
| 37 tRNA <sup>Thr</sup>       | 15534           | 15607 | 74        |       |      |        |
| 38 tRNA <sup>Pro</sup>       | 15617           | 15684 | 68        |       |      | L      |
| 39 Control Region            | 15685           | 17514 | 1830      |       |      |        |

phylogenetic trees was obtained using the NJ, MP, and Bayesian methods with the rabbit fish *Chimaera monstrosa* as an outgroup. This topology is shown in Fig. 4 with bootstrap values. Phylogenetic analysis revealed that fish from the order Myliobatiformes were well separated according to taxonomic levels

such as family, genus, and species. This appears to support the topology obtained in previous molecular studies (Dunn *et al.*, 2003). Conflicting results regarding the phylogenetic relationships among Urolophidae, Plesiobatidae, and Hexatrygonidae have been reported by previous morphological studies.



**Fig. 4** Phylogenetic relationships among myliobatoidei families. The 50%-majority rule consensus of post-burn-in sampled trees from Bayesian inference analysis based on the 12S rRNA data set under GTR + I + G model is shown. Branch support values estimated by bootstrap pseudo-replicates in Bayesian inference, maximum parsimony and neighbor-joining, respectively, are shown above each branch. (-) indicates that either bootstrap value or Bayesian posterior probabilities below 50% in the analysis. Nodes with either BP or BPP below 50% are not numbered.

Nishida (1990) placed *Plesiobatis* and *Hexatrygon* as two basal lineages of Myliobatiformes, whereas McEachran *et al.* (1996) placed *Hexatrygon*, *Plesiobatis*, and *Urolophus* as a basal lineage. Based on the present study, *Plesiobatis* and *Hexatrygon* form a sister group, which supports the hypothesis proposed by Nishida (1990). Conversely, most morphological studies indicated a closer relationship between *Urolophus* and *Plesiobatis* (Nishida, 1990; McEachran *et al.*, 1996; McEachran and Aschliman, 2004). However, the sister relationship between *Urolophus* and *Plesiobatis* was not supported by the present study. Also in this study, the families Plesiobatidae and Urolophidae were separated from each other. The family Urolophidae was more closely related to Dasyatidae than to Plesiobatidae or Hexatrygonidae. This was in accordance with the hypothesis proposed by Nelson (2006). The molecular phylogeny of Myliobatoidei presented here is only

tentative because several important genera were not included in our phylogenetic analyses. Moreover, it is important to clarify the placement of *Plesiobatis* within Myliobatiformes. Because no future resolution on the relative position of *Plesiobatis* was achieved, future work based on more complete mitochondrial genomes of Myliobatiformes will likely yield better resolved phylogenies.

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## APPENDIX

The 12 species of Myliobatoidei used for phylogenetic analysis in this study were as follows: *Urolophus aurantiacus* (AF448028); *Platyrhina sinensis* (AF448004); *Hexatrygon taiwanensis* (AF447995); *Aetoplatea zonura* (AF447986); *Rhinoptera javanica* (AF448019); *Manta birostris* (AF448000); *Mobula formosana* (AF448001); *Dasyatis kuhlii* (AF447991); *Dasyatis thetidis* (AF447993); *Urogymnus asperrimus* (AF448027); *Himantura gerrardi* (AF447996); *Himantura uarnak* (AF447997); *Squalus acanthias* (Y18134); *Chimaera monstrosa* (AJ310140).

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## 達氏深水尾魴 (*Plesiobatis daviesi*) 之完整粒線體 DNA 序列分析

蕭聖代<sup>1</sup>·徐基新<sup>2</sup>·劉燈城<sup>3</sup>·陳義雄<sup>4\*</sup>

<sup>1</sup>行政院農業委員會水產試驗所海洋漁業組

<sup>2</sup>國立中山大學海洋生物科技暨資源學系

<sup>3</sup>行政院農業委員會水產試驗所

<sup>4</sup>國立台灣海洋大學海洋生物研究所

### 摘 要

本研究利用 Long PCR 及 primer walking 的方式將達氏深水尾魴完整粒線體 DNA 完成定序，並分析基因排序與組成。達氏深水尾魴完整粒線體 DNA 全長 17514 個鹼基對，共由 37 個基因所組成，包含 2 個核糖核酸基因 (rns、rnl)、13 個蛋白質基因 (atp6、atp8、cox1-3、cob、nad1-6、nad4L) 及 22 個傳遞者核酸基因 (tRNA)，與其他脊椎動物相似。達氏深水尾魴與其他 7 種軟骨魚類的完整粒線體基因組的排序上並無明顯不同，但是達氏深水尾魴其介於 tRNA<sup>Pro</sup> 和 tRNA<sup>Phe</sup> 之間的主要非編碼區，又稱 D-loop，是目前已知的軟骨魚序列中最長的，共 1830 個鹼基對。而這段序列包含了兩段 47 個鹼基對的重複性片段，以及兩段具有保守性特徵的片段。建構其親緣關係樹後發現，達氏深水魴在演化上是與六鰓魴科為姊妹群，並且屬於燕魴亞目的基群。

關鍵詞：粒線體基因組，Long PCR，深水尾魴 (*Urotrygon daviesi*)，達氏深水尾魴 (*Plesiobatis daviesi*)